

1 COVID-19 Mass Spectrometry Coalition – Action to Accelerate Diagnostics and Treatment

2 The novel coronavirus disease 2019 (COVID-19), caused by the SARS-CoV-2 virus, was declared a  
3 pandemic by the World Health Organisation on 12<sup>th</sup> March 2020 following its emergence in Wuhan  
4 China.<sup>1</sup> As of the 4<sup>th</sup> April 2020 there were over 1.2M confirmed cases of COVID-19 in 175 countries,  
5 with over 65,000 fatalities.<sup>2</sup> SARS-CoV-2 is one of three new pathogenic coronaviruses which have  
6 jumped from animal to human hosts over the past 20 years, and the current pandemic sends warning  
7 signs about need for preparedness and associated research.

8 Today's ability to undertake rapid and comprehensive genetic sequencing has shed light on the virus'  
9 origin from animal reservoirs. This has been matched by the rapid development and implementation  
10 of PCR tests to determine the presence of viral RNA in infected individuals, as well as the ongoing  
11 deployment of serological assays to detect antibodies that recognize the SARS-CoV-2 antigen<sup>3,4</sup>. The  
12 successful deployment of these tests relies on accurate descriptors of the viral antigens and of the  
13 host response. All tests require comprehensive validation and must be done on many clinical samples  
14 with community-wide standards for the required reagents as well as for the adopted methodologies.

15 Despite the considerable insight gained into SARS-CoV-2 at the genetic level, structural information  
16 continues to emerge and, as it does, our understanding of molecular descriptors that contribute to  
17 disease progression become less well defined, especially when we consider spread across different  
18 populations.<sup>3,5</sup> The provision of rapid, precise and reproducible diagnostic information that  
19 complements genomic information, at the proteomic and metabolomic level through the power of  
20 mass spectrometry therefore needs to be deployed now in an internationally coordinated effort to  
21 accelerate our understanding of COVID-19 disease.

22 The questions that can be answered by mass spectrometry-based analysis broadly fall into three  
23 categories. The first concerns profiling the host response, where serological testing relies on  
24 immunochemical approaches to understand signalling and immune responses during various stages  
25 of infection, and correlate prognosis with disease severity. Other biomarkers of host response will  
26 further our understanding of disease mechanisms and the susceptibility of certain groups.<sup>6</sup> In this  
27 context, the most valuable markers will be those indicating the transition from a beneficial immune  
28 response to a harmful inflammatory response and respiratory distress. Such data will allow us to  
29 screen high-risk populations, recognize high risk patients, track disease progression and identify  
30 sources of vulnerability that will help treat and prevent future coronavirus pandemics.

31 The second question, and arguably the most urgent, concerns the SARS-CoV-2 viral spike glycoprotein,  
32 as it is not only key for host-cell attachment but also as a target for neutralizing antibodies elicited  
33 through vaccination. Whilst RNA sequencing is extraordinary informative with regards to viral  
34 mutation or adaptation via immune selective pressure, it cannot inform on a critical feature among  
35 enveloped viruses, specifically viral spike glycosylation.

36 Many viruses exploit the glycosylation machinery, using host glycans for immune evasion, for example  
37 HIV-1 hiding immunological epitopes behind a "glycan shield". The functional role of SARS-CoV-2 spike  
38 glycans, of which there are 66 per trimer<sup>7</sup>, is presently undetermined and yet is key for structure-  
39 based vaccine design, receptor (ACE2) binding and associated conformational dynamics which  
40 influence binding efficacy.<sup>8</sup> Measuring spike glycosylation and plasticity is crucial for the production  
41 of vaccine candidates which mimic circulating viruses and for drug development efforts to block  
42 receptor attachment and subsequent infection.

43 The final area, which is critical for policy decision and guidance, concerns *in-situ* analyses. It is still  
44 unclear how stable and infectious is SARS-CoV-2 on surfaces, and underlines the need for a robust

45 diagnostic for viral material as well as for RNA. SARS-CoV-2 RNA has been detected on various surfaces  
46 several weeks after they were last touched<sup>9</sup>, and also in sewage<sup>10</sup>, whether either source also has an  
47 infectious protein complement is unknown. Identification of the viral antigen and other biomarkers  
48 with mass spectrometry from various contaminated sources at defined time points, could fill this gap  
49 in knowledge and help to adopt caution and protective measures.

50 This collective mass spectrometry effort will provide molecular level information of SARS-CoV-2 in the  
51 human host. We propose an international and a comprehensive mass spectrometry-based omics and  
52 native approach that will reveal pathophysiological and structural information to treat COVID-19.  
53 Working to community standards<sup>11,12</sup>, we will report all datasets, to complement genomic data. We  
54 need to act now by repurposing mass spectrometry resources in university labs and research institutes  
55 across the world that are routinely used for disease diagnosis, characterisation of complex  
56 biopharmaceuticals and drug discovery, to help tackle this and potential future pandemics. It is  
57 essential that mass spectrometry is now used collectively to quantify the viral antigens used in  
58 serological diagnostics and protein-based antigens.

59 Working with clinical colleagues, our efforts will allow the sharing of methods for sample collection,  
60 processing and data formatting. Open datasets will allow the computational community access to  
61 valuable information to determine the mechanisms behind antigen response and inform vaccine  
62 development, *in-silico* and *in-vitro* antiviral drug development. As countries across the world ramp up  
63 testing along with widespread sampling to confirm SARS-CoV-2 exposure and measure immunity,  
64 mass spectrometry has a significant role to play, and through these collaborative actions we will  
65 benefit all.

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67 Fernando Corrales, Andrea Sinz and Perdita Barran, on behalf of the COVID-19 MS Coalition

- 68 1 X. Yang, Y. Yu, J. Xu, H. Shu, J. Xia, H. Liu, Y. Wu, L. Zhang, Z. Yu, M. Fang, T. Yu, Y. Wang, S.  
69 Pan, X. Zou, S. Yuan and Y. Shang, *Lancet Respir. Med.*, , DOI:10.1016/S2213-2600(20)30079-  
70 5.
- 71 2 COVID-19 Map - Johns Hopkins Coronavirus Resource Center,  
72 <https://coronavirus.jhu.edu/map.html>, (accessed 5 April 2020).
- 73 3 F. Amanat, T. Nguyen, V. Chromikova, S. Strohmeier, D. Stadlbauer, A. Javier, K. Jiang, G.  
74 Asthagiri-Arunkumar, J. Polanco, M. Bermudez-Gonzalez, D. Caplivski, A. Cheng, K.  
75 Kedzierska, O. Vapalahti, J. Hepojoki, V. Simon and F. Krammer, *medRxiv*, 2020,  
76 2020.03.17.20037713.
- 77 4 R. Wölfel, V. M. Corman, W. Guggemos, M. Seilmaier, S. Zange, M. A. Müller, D. Niemeyer, T.  
78 C. Jones, P. Vollmar, C. Rothe, M. Hoelscher, T. Bleicker, S. Brünink, J. Schneider, R. Ehmann,  
79 K. Zwirgmaier, C. Drosten and C. Wendtner, *Nature*, 2020, 1–10.
- 80 5 Q. Long, H. Deng, J. Chen, J. Hu, B. Liu, P. Liao, Y. Lin, L. Yu, Z. Mo, Y. Xu, F. Gong, G. Wu, X.  
81 Zhang, Y. Chen, Z. Li, K. Wang, X. Zhang, W. Tian, C. Niu, Q. Yang, J. Xiang, H. Du, H. Liu, C.  
82 Lang, X. Luo, S. Wu, X. Cui, Z. Zhou, J. Wang, C. Xue, X. Li, L. Wang, X. Tang, Y. Zhang, J. Qiu, X.  
83 Liu, J. Li, D. Zhang, F. Zhang, X. Cai, D. Wang, Y. Hu, J. Ren, N. Tang, P. Liu, Q. Li and A. Huang,  
84 *medRxiv*, 2020, 2020.03.18.20038018.
- 85 6 J. E. Kyle, K. E. Burnum-Johnson, J. P. Wendler, A. J. Einfeld, P. J. Halfmann, T. Watanabe, F.  
86 Sahr, R. D. Smith, Y. Kawaoka, K. M. Waters and T. O. Metz, *Proc. Natl. Acad. Sci. U. S. A.*,  
87 2019, **116**, 3919–3928.
- 88 7 Y. Watanabe, J. D. Allen, D. Wrapp, J. S. McLellan and M. Crispin, *bioRxiv*, 2020,

89 2020.03.26.010322.

90 8 A. C. Walls, Y.-J. Park, M. A. Tortorici, A. Wall, A. T. McGuire and D. Veessler, *Cell*, ,  
91 DOI:10.1016/j.cell.2020.02.058.

92 9 L. F. Moriarty et al. MMWR Morb Mortal Wkly Rep. 2020 Mar 27;69(12):347-352. DOI:  
93 10.15585/mmwr.mm6912e3.

94 10 G. Medema, L. Heijnen, G. Elsinga, R. Italiaander and A. Brouwer. medRxiv DOI:  
95 10.1101/2020.03.29.20045880

96 11 HUPO - Proteomics Standards Initiative, [https://www.hupo.org/Proteomics-Standards-](https://www.hupo.org/Proteomics-Standards-Initiative)  
97 Initiative, (accessed 5 April 2020).

98 12 The Metabolomics Standards Initiative (MSI) and Core Information for Metabolomics  
99 Reporting (CIMR) | <http://cosmos-fp7.eu>, <http://cosmos-fp7.eu/msi.html>, (accessed 5 April  
100 2020).

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